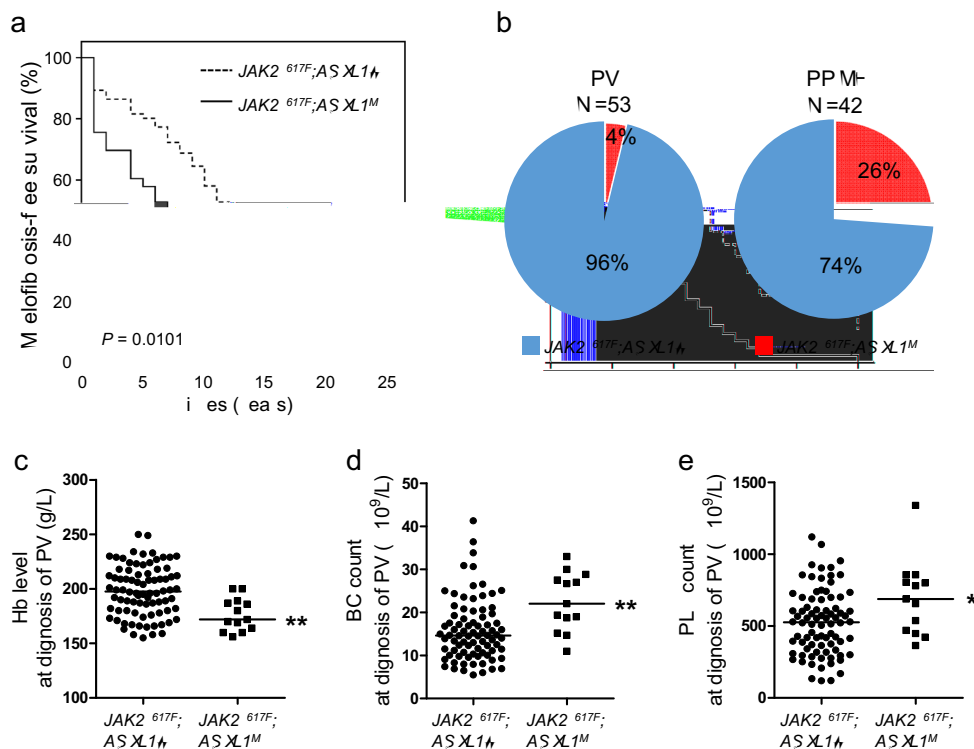




**Fig. 1** *ASXL1* alteration cooperates with *JAK2*<sup>V617F</sup> to accelerate MPN progression in PV patients. **a** Myelofibrosis-free survival analysis of *JAK2*<sup>V617F</sup> PV patients according to the *ASXL1* mutational status from a Cox regression model. *ASXL1*<sup>WT</sup>: *ASXL1*<sup>wild-type</sup> ( $n = 82$ ). *ASXL1*<sup>MUT</sup>: *ASXL1*<sup>mutated</sup> ( $n = 13$ ). Log-rank test was used for survival statistics. **b** Among the 95 PV patients with *JAK2*<sup>V617F</sup> mutations, the proportion of *ASXL1* mutations is higher in PPMF (26%) than in PV patients without MF (4%). **c–e** PB counts of Hb, WBC, and PLT in *JAK2*<sup>V617F</sup> PV patients with WT *ASXL1* or mutated *ASXL1*. \* $p < 0.05$ ; \*\* $p < 0.01$

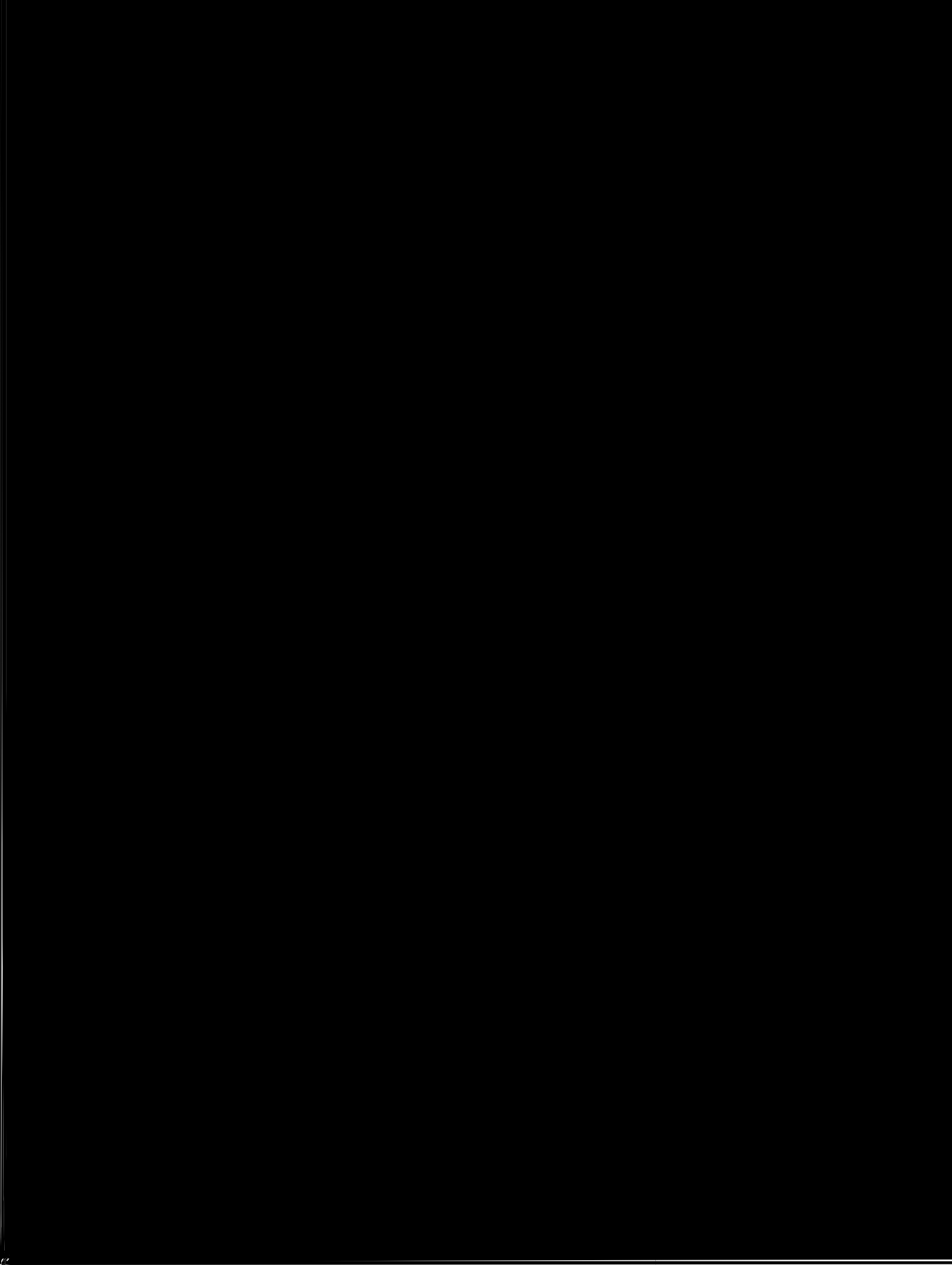


analyses including necropsy, histology, and flow cytometry on peripheral blood (PB), bone marrow (BM), and spleen. Fibrosis was assessed at each specified time points. We found that both *JAK2*<sup>V617F</sup>;*Asxl1*<sup>+/-</sup> and *JAK2*<sup>V617F</sup> mice developed progressive MPN, including PV, ET, and MF (Supplementary Figure S1B). Notably, 5 (26%) of the 19 *JAK2*<sup>V617F</sup>;*Asxl1*<sup>+/-</sup> mice developed MF at 2–6 months of age, which was much earlier than *JAK2*<sup>V617F</sup> mice (1 (6%) of the 18 mice develop MF at 6 months of age) (Fig. 2a). Additionally, 3 (12%) of the 26 *JAK2*<sup>V617F</sup>;*Asxl1*<sup>+/-</sup> mice, but not the littermate controls, progressed/transformed to secondary acute myeloid leukemia (Fig. 2a, Supplementary Figure S1B–C).

The average numbers of WBC, neutrophil, and PLT were significantly higher in the PB of *JAK2*<sup>V617F</sup>;*Asxl1*<sup>+/-</sup> mice compared with the wild-type (WT) group at the age of 2–3 months. In contrast, there was no significant difference in the WBC and neutrophil counts in the *JAK2*<sup>V617F</sup> mice at the age of 2–3 months, and from 4–6 months of age, the *JAK2*<sup>V617F</sup> mice started to exhibit higher counts of WBC and neutrophil (Fig. 2b–d). The Hb levels in *JAK2*<sup>V617F</sup>;*Asxl1*<sup>+/-</sup> mice with MF were the lowest among all the groups of mice at the age of 8–10 months (Supplementary Figure S2A). The *JAK2*<sup>V617F</sup>;*Asxl1*<sup>+/-</sup> mice exhibited splenomegaly (Supplementary Figure S2B). The histologic analysis of the femur sections from *JAK2*<sup>V617F</sup>;*Asxl1*<sup>+/-</sup> mice displayed megakaryocytic hyperplasia (Fig. 2e). Spleen sections from *JAK2*<sup>V617F</sup>;*Asxl1*<sup>+/-</sup> mice showed a disrupted splenic architecture and prominent

megakaryocytes and myeloid precursors (Fig. 2e). Reticulin staining of femur sections revealed extensive fibrosis in the BM of *JAK2*<sup>V617F</sup>;*Asxl1*<sup>+/-</sup> mice at the age of 3 months (Fig. 2e). Consistently, flow cytometric analysis demonstrated that *JAK2*<sup>V617F</sup>;*Asxl1*<sup>+/-</sup> mice had a significant expansion of erythroid precursors (Ter119<sup>+</sup>CD71<sup>+</sup>) in the BM and spleen compared with the *JAK2*<sup>V617F</sup> and WT groups (Supplementary Figure S2C). The PB smear of *JAK2*<sup>V617F</sup>;*Asxl1*<sup>+/-</sup> mice contained more bluish red blood cells compared with WT PB smear, suggesting an impaired red blood cell differentiation (Supplementary Figure S2D). Interestingly, the frequency of CD41<sup>+</sup>CD61<sup>+</sup> megakaryocytic precursors was significantly increased in the BM of *JAK2*<sup>V617F</sup>;*Asxl1*<sup>+/-</sup> mice by flow cytometric analysis compared with WT and *JAK2*<sup>V617F</sup> mice (Supplementary Figure S2E). These data demonstrate that concurrent haploinsufficiency of *Asxl1* and *JAK2*<sup>V617F</sup> enhances megakaryopoiesis and increases erythroid precursors in the BM and spleen, which may accelerate MF development in vivo.

To determine the effect of *Asxl1* alteration on *JAK2*<sup>V617F</sup> HSC/HPCs, we performed flow cytometric analyses and found that the frequencies of short-term (ST)-HSC and megakaryocyte/erythroid progenitor (MEP) were significantly increased in the BM of *JAK2*<sup>V617F</sup>;*Asxl1*<sup>+/-</sup> mice compared with those in WT mice, while the frequency of multipotent hematopoietic progenitors (MPP) was significantly decreased in the BM of *JAK2*<sup>V617F</sup>;*Asxl1*<sup>+/-</sup> mice compared with that in WT mice (Supplementary Figure S3A–B). Colony-forming unit (CFU) assays



revealed the total colony number and replating capacity in the BM cells of  $JAK2^{V617F};Asx1I^{+/-}$  mice was significantly higher compared with those in the WT group of mice (Supplementary Figure S3C–D). Furthermore, we observed the frequencies of burst-forming unit-erythroid (BFU-E), CFU-erythroid (CFU-E), and CFU-megakaryocyte (CFU-MK) colonies were significantly increased in response to a series of doses of erythropoietin (EPO) in the spleen and BM cells from  $JAK2^{V617F};Asx1I^{+/-}$  mice (Fig. 2f, Supplementary Figure S3E). Notably, EPO-independent BFU-E and CFU-E formation, a hallmark feature of PV [15], were also higher in the spleen and BM cells from  $JAK2^{V617F};Asx1I^{+/-}$  mice compared with other groups of mice (Fig. 2f, Supplementary Figure S3E). Thus *Asx1I* alteration cooperates with *JAK2V617F* mutation leading to biased lineage skewing, favoring erythroid and megakaryocytic differentiation.

We have reported that leukemic transformation in MDS/MPN can occur in the aged *Asx1I*<sup>+/-</sup> mice (>16 month old) [14]. In the current study, we found that three  $JAK2^{V617F};Asx1I^{+/-}$  mice developed myeloid leukemia at the age of 6–8 months, which is much earlier than that found in *Asx1I*<sup>+/-</sup> mice. Two of the  $JAK2^{V617F};Asx1I^{+/-}$  mice also had intestinal myeloid sarcoma, which were verified by histology and flow cytometric analysis (Supplementary Figure S4A–C). The moribund leukemic  $JAK2^{V617F};Asx1I^{+/-}$  mice had blast cells in PB, >20% blast cells in BM, and splenomegaly (Supplementary Figure S4D–G). The histologic analyses of femur sections revealed an increase of megakaryocytes and a decrease in erythroid islands of these  $JAK2^{V617F};Asx1I^{+/-}$  mice but not in any other groups of mice. The spleen sections of  $JAK2^{V617F};Asx1I^{+/-}$  mice showed a disrupted architecture with an increased proportion of myeloid cells. Reticulin staining showed extensive BM fibrosis in  $JAK2^{V617F};Asx1I^{+/-}$  mice (Supplementary Figure S4H). These data indicate that *Asx1I* alteration cooperates with *JAK2V617F* mutation to accelerate myeloid leukemic transformation.

In summary, PV patients with co-mutations of *ASXL1* and *JAK2V617F* had a poor MF-free survival. Likewise, *Asx1I* loss accelerates MF in *JAK2V617F*-driven MPN in mice.  $JAK2^{V617F};Asx1I^{+/-}$  mice induces megakaryocytic hyperplasia and can transform to myeloid leukemia. Future studies using the *Asx1I* and  $JAK2^{V617F}$  co-mutated mice to further investigate the cooperative effect between *ASXL1* mutant and *JAK2V617F* in the progression of myeloid malignancies are warranted.

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## Compliance with ethical standards

**Conflict of interest** The authors declare that they have no conflict of interest.

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